

MSⁿ spectral tree databases: annotation and identification of plant metabolites

Justin J.J. van der Hoof^{1,2,3}, Miguel Rojas-Cherto³, Thomas Hankemeier³, Michael van Vliet³, Theo Reijmers³, Robert Hall^{1,2,3} & Ric de Vos^{1,2,3}

¹ Plant Research International, Wageningen-UR, Wageningen, The Netherlands

² Centre for BioSystems Genomics, Wageningen, The Netherlands

³ Netherlands Metabolomics Centre, Leiden, The Netherlands

ric.devos@wur.nl



Introduction and Aim

MSⁿ tools for annotation and identification of metabolites detected in LC-MS profiling are under development. Since MSⁿ spectral trees are robust,^{1,2} MSⁿ databases and dedicated software able to process and handle MSⁿ data are key to efficiently implement MSⁿ in the metabolomics pipeline.

Here we examine the use of high mass resolution MSⁿ data as input for a plant spectral tree database that can be populated, visualized, and queried, using recently developed MetiTree software³, encompassing MSⁿ processing tools like MEF⁴ and a spectral tree comparison algorithm.⁵

MSⁿ data generation

High mass resolution MSⁿ fragmentation data was obtained by using a LC-PDA-LTQ MS-Orbitrap FTMS (Thermo) combined with a Nanomate fraction collector / injector (Advion) with a chip-based nanospray source (Figure 1).



Figure 1: Schematic overview of MSⁿ spectral tree data generation.

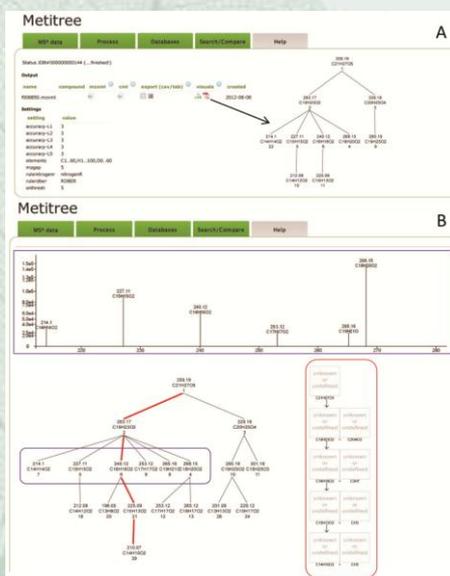


Figure 2: MetiTree software tool. A) MetiTree output and visualization of a processed spectral tree. B) Example of a tree of fragments (left) assigned with the corresponding elemental formulas of fragments and losses (right) of a fragmentation path (marked in red) as obtained from a C₂₁H₂₈O₅ diterpene related compound. The MS³ spectrum (marked in purple) is shown on top.

MSⁿ spectral tree processing

The MetiTree software assigns elemental formulas to both the parent ion of a fragmented metabolite and its daughter ions.⁴ Using the hierarchical relation of the fragment ions, the elemental formulas are now the nodes of the spectral trees (Figure 2). Upon processing of the raw MSⁿ files, also chemical and electronic noise peaks are removed, which facilitates comparison between spectral trees. As proof of principle, a spectral tree database was created and populated with 15 (phenolic) diterpenes and related structures present in *Rosmarinus officinalis* trichomes for querying unknown structures (Figure 3).

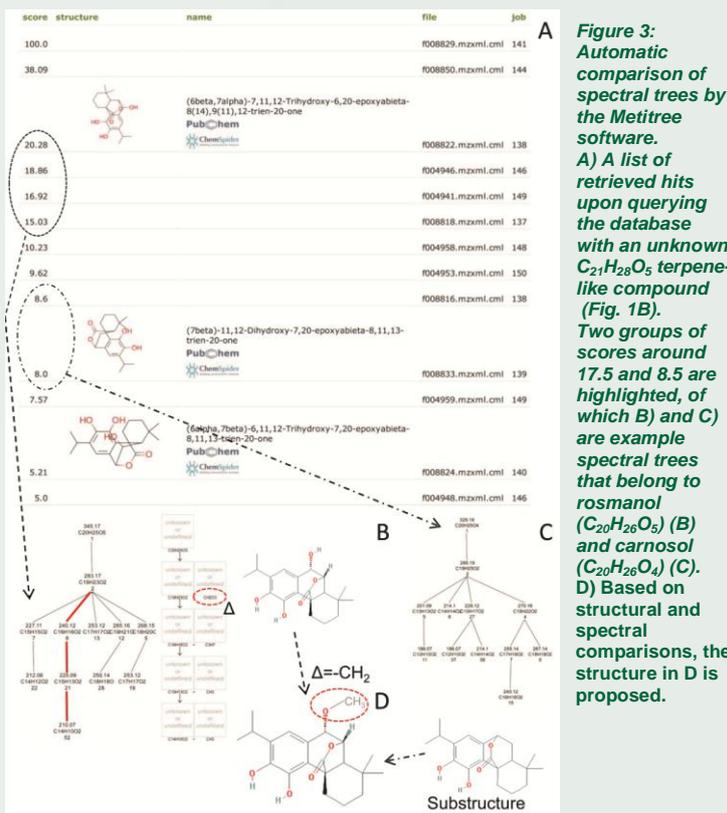


Figure 3: Automatic comparison of spectral trees by the MetiTree software. A) A list of retrieved hits upon querying the database with an unknown C₂₁H₂₈O₅ terpene-like compound (Fig. 1B). Two groups of scores around 17.5 and 8.5 are highlighted, of which B) and C) are example spectral trees that belong to rosmanol (C₂₀H₂₆O₅) (B) and carnosol (C₂₀H₂₆O₄) (C). D) Based on structural and spectral comparisons, the structure in D is proposed.

Conclusions

- Robust MSⁿ data of plant metabolites can be used to populate spectral tree databases.
- Comparing spectral trees from yet unknown compounds to already fragmented metabolites using MSⁿ databases facilitates structural elucidation.

References

1. Van der Hoof, J. J. J. et al. (2011). "Polyphenol identification based on systematic and robust high-resolution accurate mass spectrometry fragmentation." *Anal. Chem.* 83(1): 409-416.
2. Van der Hoof, J. J. J. et al. (2012). "Spectral trees as a robust annotation tool in LC-MS based metabolomics." *Metabolomics* 8(4): 691-703.
3. Rojas-Cherto, M. et al. (2012). "MetiTree: a web application to organize and process high resolution multi-stage mass spectrometry metabolomics data." *Bioinformatics*.
4. Rojas-Cherto, M. et al. (2011). "Elemental composition determination based on MSⁿ." *Bioinformatics* 27(17): 2376-2383.
5. Rojas-Cherto, M. et al. (2012). "Metabolite identification using automated comparison of high resolution MSⁿ spectral trees." *Anal. Chem.* 84(13): 5524-5534.